

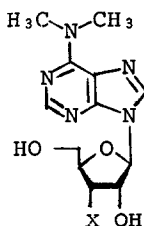
PERMANGANATE OXIDATION OF N⁶,N⁶,8-TRISUBSTITUTED-2',3',5'-
TRI-O-ACETYLADENOSINES

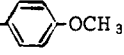

Tetsuo Kato, Shuichi Ogawa^{*a)}, and Isoo Ito^{b)}

- a) Chemical Research Division, General Laboratories, Arakawa
Chotaro & Co., 1, Hananoki, Kusakabe, Inazawa, Aichi 492, Japan
b) Faculty of Pharmaceutical Sciences, Nagoya City University,
Tanabe-dori, Mizuho-ku, Nagoya 467, Japan

Summary: Oxidation of 2',3',5'-tri-O-acetyl derivatives of N⁶,N⁶-dialkyladenosines (3a-g) with KMnO₄ in 50% AcOH gave both the mono (5a-g) and didealkyl derivatives (6a-c); it was conclusively proved that one and two methylene groups of the title nucleosides (3a-g) in the α position to the exocyclic nitrogen atom were simultaneously oxidized.

N⁶,N⁶-Dimethyladenosine (1a) is found as the component of RNA¹⁾ or as antibiotic puromycin (1b)²⁾. The nucleoside (1a) occurs as a part of 16s and 18s ribosomal RNA which is believed to be responsible for the binding of antibiotic kasugamycin³⁾. Very recently, one of authors has reported the synthesis of such a "reduced" analogue of puromycin, compound (1c)⁴⁾. Continuously, we attempted to synthesize 8-substituted-N⁶,N⁶-dimethyladenosines, because it is of interest to know the structure-activity relationships between 3' and 8-substituted derivatives.



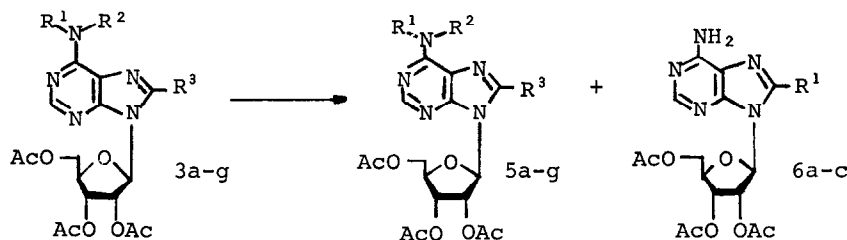
- 1a) X = OH
1b) X = NHCOCHCH₂-
1c) X = NHCH₂CHCH₂-
1d) X = NH₂

In general, 8-bromo- N^6,N^6 -dimethyladenosine(2)⁵⁾ is useful intermediate for the preparation of 8-halo, 8-amino, and 8-thio- N^6,N^6 -dimethyladenosine. Attempts to displace the 8-bromo group with inactive nucleophiles such as secondary amines or amino acids have been unsuccessful. A methylsulfonyl group is known to be a good leaving group in the reaction with nucleophiles in nucleosides⁶⁾. Thus, we attempted to synthesize 8-methylsulfonyl- N^6,N^6 -dimethyl-2',3',5'-tri-O-acetyladenosine(4) from 8-methylthio- N^6,N^6 -dimethyl-2',3',5'-tri-O-acetyladenosine(3g)⁷⁾ by oxidation.

We found the mono and didemethylation of N^6,N^6 -dimethyl group under the condition of $KMnO_4$ -50% AcOH, and 3g was converted to N^6 -methyl-8-methylsulfonyl-2',3',5'-tri-O-acetyladenosine(5g) and 8-methylsulfonyl-2',3',5'-tri-O-acetyladenosine(6c)⁶⁾. Puromycin(lb) is converted in vivo to the highly nephrotoxic amino-nucleoside(ld), which is then selectively monodemethylated and phosphorylated to the corresponding 5'-phosphate⁸⁾. Recently, Žemlička, et.al.,⁹⁾ have reported a selective N^6 -monodemethylation sequence with RuO_4 as a nonenzymatic model.

We now wish to report the result of the oxidation of a series of N^6,N^6 -8-trisubstituted-2',3',5'-tri-O-acetyladenosines(3a-g) with $KMnO_4$ in a polar solvent (50% AcOH), and the difference in the reactivities between $KMnO_4$ and RuO_4 as a oxidizing reagent.

Table I. The results of oxidation of N^6,N^6 -8-trisubstituted-2',3',5'-tri-O-acetyladenosines(3a-g) with $KMnO_4$ under various conditions¹⁰⁾



Starting Material				Amount of $KMnO_4$ (equ.)	Temp (°C)	Time (hr)	Yield (%)			Products		Yield (%)		
No.	R^1	R^2	R^3				No.	R^1	R^2	R^3	No.		R^1	
3a	CH ₃	CH ₃	H	1	r.t.	1.5	5a	CH ₃	H	H	26.3	6a	H	none
3a	CH ₃	CH ₃	H	3	40	1	5a	CH ₃	H	H	32.3	6a	H	48.1
3b	C ₂ H ₅	C ₂ H ₅	H	3	r.t.	0.5	5b	C ₂ H ₅	COCH ₃	H	42.5	6a	H	37.3
3b	C ₂ H ₅	C ₂ H ₅	H	3	r.t.	1	5b	C ₂ H ₅	COCH ₃	H	38.9	6a	H	43.7
5b	C ₂ H ₅	COCH ₃	H	3	r.t.	0.5						6a	H	83.2
3c	CH ₂ -	CH ₂ -	H	3	r.t.	0.67	5c	CH ₂ -	H	H	23.3	6a	H	28.4
3d	(CH ₂) ₄	H	H	3	40	1	5d	(CH ₂) ₃ CO	H	H	48.2	6a	H	4.3
3e	(CH ₂) ₅	H	H	3	r.t.	0.17	5e	(CH ₂) ₄ COOH	H	H	31.1	6a	H	38.2
3f	CH ₃	CH ₃	Br	3	40	0.17	5f	CH ₃	H	Br	36.0	6b	Br	31.3
3g	CH ₃	CH ₃	SCH ₃	3	4	1.5	5g	CH ₃	H	SOCH ₃	52.3	6c	SO ₂ CH ₃	6.5

The results of the oxidation of the title series of nucleosides (3a-g) are summarized in Table I. N⁶,N⁶-Dimethyl-2',3',5'-tri-O-acetyladenosine (3a) was oxidized with 1 equivalent of KMnO₄ at room temperature to give the N⁶-methyl derivative (5a)⁹⁾. While 3a was oxidized with 3 equivalents of KMnO₄ at 40°C to give the didemethylated compound, 2',3',5'-tri-O-acetyladenosine (6a)¹¹⁾ along with 5a. The properties of 5a and 6a were identical with those of authentic samples by PMR spectra, UV spectra, CD spectra, TLC of silicagel, and PPC. In addition, deacetylation of 5a and 6a with methanolic ammonia gave N⁶-methyladenosine¹²⁾ and adenosine, respectively, whose properties were in accord with those of an authentic sample.

N⁶,N⁶-Diethyl-2',3',5'-tri-O-acetyladenosine (3b)⁹⁾ afforded the N⁶-acetyl-N⁶-ethyl derivative (5b)⁹⁾ and 6a. Compound (5b) was deacetylated with methanolic ammonia to N⁶-ethyladenosine¹³⁾, identical with an authentic sample. Further oxidation of 5b did not give the intermediate, N⁶,N⁶,2',3',5'-O-pentaacetyladenosine, but afforded 6a readily. The isolation of 5b suggests the reaction mechanism of 3a to 5a proceeds as follows; 5b might be formed in some preceding step, and then, decomposition of the carbinolamine or formylamine intermediate, probably took place.

N⁶,N⁶-Dibenzyl-2',3',5'-tri-O-acetyladenosine (3c)⁹⁾ proved to be less reactive in comparison with 3a and 3b, and N⁶-benzyl derivative (5c), and 6a were obtained in 23% and 25% yield, respectively. It seemed that the intermediate, N⁶-benzoyl-N⁶-benzyl derivative was unstable under the acidic condition, and the phenyl group was rapidly oxidized with KMnO₄. The structure of 5c was confirmed by deacetylation with ammonia in methanol to the known N⁶-benzyladenosine¹⁴⁾.

As the oxidation of aliphatic amines with KMnO₄ was successful, we next examined the oxidation of some purine derivatives substituted at position 6 with a cyclic amine. It was thought that such a reaction would lead to novel 6-substituted purine derivatives which were of interest biologically. The pyrrolidino derivative (3d) gave the corresponding lactam (5d) and 6a by the oxidation with KMnO₄, under the condition described above. The corroboration of the structure came from the PMR spectrum of 5d similar to the corresponding tribenzoyl derivative⁹⁾. The piperidino derivative (3e) gave on oxidation with KMnO₄, N-(2',3',5'-tri-O-acetyladenosin-6-yl)valeric acid (5e), and 6a. The structure of 5e was confirmed by deacetylation with ammonia in methanol to N-(adenosin-6-yl)valeric acid which was identical with a sample prepared by alkylation of 6-chloroinosine¹⁵⁾ with 5-amino-*n*-valeric acid in dimethylformamide.

Oxidation of 8-bromo-N⁶,N⁶-dimethyl-2',3',5'-tri-O-acetyladenosine (3f) gave the similar products, 8-bromo-N⁶-methyl (5f), and 8-bromo (6b)¹⁶⁾ derivative, to the above cases. Similarly, compound (3g) was converted to 5g and 6c. The PMR spectrum of 5g showed that thiomethyl signal of 3g at δ 2.71 was deshielded to δ 3.41 based on methylsulfonyl group.

Žemlička, et.al.,⁹⁾ have reported that selectivity of oxidation of N⁶,N⁶-

dialkyl-nucleosides with RuO_4 can be explained in terms of electronic effects of N^6 -alkyl group, the pyrimidine portion of the purine ring, and the carbonyl function in the reaction products. We, however, found both the mono and didealkylation of N^6, N^6 -dialkyladenosines on oxidation with KMnO_4 ; it was conclusively proved that one and two methylene groups of the title nucleoside in the α position to the exocyclic nitrogen atom were simultaneously oxidized. The fact might be suggested that KMnO_4 is more powerful than RuO_4 in the oxidation of the N-alkyl group, and each alkyl group in N^6, N^6 -dialkyl derivatives is equivalent.

It will be possible to utilize KMnO_4 as a removal of N^6 -blocking group under the reaction condition in regard as the studies on cyclonucleosides, nucleotides, and oligonucleotides. Oxidation of N^6 -monoalkyl derivatives, and other oxidizing reagents are being currently explored in our laboratory.

Acknowledgement

The authors wish to express their gratitude to Dr.T.Ueda for mass spectral measurements, to the microanalytical center of the faculty for elemental analyses, to Mrs.T.Kumagai for PMR spectral measurements, and to Mr.K.Miyazaki for his technical assistance.

References

- 1) D.B.Dunn, *Biochim.Biophys.Acta.*, 34, 286(1959).
- 2) B.R.Baker, R.E.Schaub, and J.H.Williams, *J.Amer.Chem.Soc.*, 77, 7(1955).
- 3) T.L.Helser, J.E.Davis, and J.E.Dahlberg, *Nature(London), New Biol.*, 233, 12(1971).
- 4) T.Kato, and J.Žemlička, *Tetrahedron Letters*, 48, 4741(1978); *J.Org.Chem.*, 45, 4006(1980).
- 5) M.Ikehara, and H.Morisawa, *Chem.Pharm.Bull.*, 19, 2593(1971).
- 6) A.Matsuda, Y.Nomoto, and T.Ueda, *Chem.Pharm.Bull.*, 27, 183(1979).
- 7) M.Ikehara, and T.Maruyama, *Chem.Pharm.Bull.*, 24, 565(1976).
- 8) H.T.Nagasawa, K.F.Swingle, and C.S.Alexander, *Biochem.Pharmacol.*, 16, 2211(1967); R.F.Derr, D.K.Loehler, C.S.Alexander, and H.T.Nagasawa, *ibid.*, 17, 265(1968).
- 9) T.Endo, and J.Žemlička, *J.Org.Chem.*, 44, 3652(1979).
- 10) AcOH(50%) was used as a solvent, except 3c was reacted in 80% AcOH.
- 11) D.B.Dunn, *Biochim.Biophys.Acta.*, 1961, 46,198.
- 12) D.B.Dunn, J.D.Smith, and P.F.Spahr, *J.Mol.Biol.*, 1960, 2, 113.
- 13) J.A.Montgomery, and H.J.Thomas, *J.Org.Chem.*, 28, 2304(1963).
- 14) R.E.Holmes, and R.K.Robins, *J.Amer.Chem.Soc.*, 86, 1242(1964).
- 15) J.Zemlicka, and J.Owens in "Nucleic Acid Chemistry-Improved and New Synthetic Procedures, Methods and Techniques", L.B.Towensend and R.S.Tipson, Eds., Wiley, New York, 1978, Part 2, p 611.
- 16) J.Žemlička, and F.Šorm, *Collect.Czech.Chem.Comm.*, 30, 1880(1965).

(Received in Japan 11 May 1981)